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Profile Of Children's Urinary Tract Infection, Evaluation Of Risk Factors And Review Of The Literature

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Abstract:

It is important to indicate the distribution and alteration of the factors of urinary tract infections in children and of the resistance ratios according to years and to probe the accompanying risk factors. In this study, the records of Microbiology Laboratory and Pediatric Clinic of Izmir Menemen State Hospital were searched retrospectively. In the course of the study, 10^5 cfu/ml and single microorganism growth were taken under evaluation in urine cultures. A total of 2669 urine cultures were evaluated; in 456 of them (17%) significant growth was detected in terms of urinary tract infection. 275 (60%) of the samples with reproduction belonged to girls and 181 (40%) of them belonged to boys. Escherichia coli was the most commonly reproduced microorganism in 64% patients. Klebsiella spp. was determinant in %16 patients, Proteus spp. in %8 patients, Enterococcus spp. in %5 patients, Pseudomonas aeruginosa in %3 patients, Staphylococcus aureus in %2 patients, Coagulase Negative Staphylococcus in %2 patients. In E.coli strains, 66% resistance was detected to ampicilline, 16% to amoxicilline clavulanate, 28% to ampicilline/sulbactam, 14% to ceftazidime, 25% to ceftriaxone, 6% to nitrofurantoine, 6% to phosphomisine, 0% to imipenem, 29% to trimethoprim sulfamethoxazole, 23% to gentamicine. Ampicillin was the antibiotic against which the highest resistance was determined by 100% for Klebsiella spp. and by 71% for Proteus spp. In the childhood urinary tract infections, more resistance to ampicillin, ampicillin/sulbactam, gentamicin and TMP-SMX, frequently preferred as the first option, was found than the others. During this study period, the resistance to meropenem and imipenem was not found in childhood group. ESBL positive Gram negative bacteria rate was 69/456 (15%). A total of 59 of them were E.coli, 10 of them were Klebsiella spp.

Today, nitrofurantoin, aminoglycosides are the antibiotics that can still be suggested in empirical treatment due to the low resistance. Yet, the continuation of the antibiotic treatments started empirically must be decided again after the culture and susceptibility test results reported by microbiology laboratory are interpreted, as suggested in guides.

Key Words: Pediatric urinary tract infections, antibiotic resistance, risk factors

Introduction

Urinary Tract Infections (UTI) are frequent clinical pictures in the first five years in childhood (1). As the factor can be bacteria, viruses or fungi can cause them (2). Both anatomic and some disorders that can happen functionally pave the way for the formation of infections in urinary tract (3). The reflux or stone in the urinary tract are examples of functional disorders that pave the way for UTI. One of the reasons that increase the prevalence in urinary tract infections in children is the deficiency of hygiene in genital region (4, 5). Also, swimming pools not being hygienic enough is another risk factor for each age group. In children with chronic constipation problem, more frequent infections can happen in urinary tract because of long accumulation of urine in the bladder (5). A total of 2669 urine culture tests of children examined in microbiology laboratory of our hospital in January 2014-November 2015 were probed for the profile of active microorganisms, their antibiotic susceptibility and determination of risk factors.

Material and method

The patients included in the study are 0-18 aged patients who applied to pediatric polyclinic of our hospital in January 2014- November 2015. A total of 803/2669 reproduction was detected in urine samples, including samples more than one taken at different times from the same patient. In the examination made considering one reproduction result of each patient, 275 of 456 patients detected reproduction in urine culture were females; 181 of them were males and 52 of them were under the age of 1; 404 of them were over the age of 1. The number of patients followed with the diagnosis of vesicoureteral reflux (VUR) was 13 (%2.85).

Clean urine samples taken as mid-stream in childhood and collected into drainage bag in infants were added into 5% sheep blood agar and Eosine Methylene Blue agar (Salubris, Turkey), and bacterial growth was evaluated after incubated for 18-24 hours at 37 °C. A positive culture was defined as follows: growth of a single urinary tract pathogen with at least > 10^5 colony-forming units (CFU)/mL in urine specimens. After the reproducing bacteria were completed by looking at gram stain, catalase, coagulase, oxidase tests through traditional methods and by using API 20E (BioMérieux, France) bacteria identification kit, antibiotic susceptibility tests were performed through disk diffusion method in compliance with the methods of "The Clinical and Laboratory Standards Institute" (CLSI). (6). Extended spectrum beta lactamase (ESBL) producing organisms were screened and confirmed according to the method suggested by the CLSI (6).

Results

A total of 2669 urine cultures were examined during the two-year following period. Reproduction took place in 456 (17%) of the samples. Of the samples with reproduction, 275 (60%) were taken from females, 181 (40%) from males. The number of reproduction according to gender are in

Table 1. The average age of the patients was 60 ± 24 months (1-180 months). A total of 52 of the cases (11%) were under 12 months age, 404 of them (89%) were over 12 months age. Of the 13 (2.9%) children with VUR, 6 were female, 7 were male patients. *E.coli* was the most commonly reproduced microorganism in 292 (64%) patients. *Klebsiella spp* was active in 72 (16%) patients, *Proteus spp* was active in 36 (8%) patients, *Enterococcus spp*. was active in 24 (5%) patients, *Staphylococcus aureus* was active in 9 (2%) patients, *Coagulase Negative Staphylococcus* was active in 10 (2%) patients. The resistance rates to antibiotics of the most common Gram positive bacteria in urine cultures were given in Table 2, Gram negative bacteria were given in Table 3.

Ampicillin resistance was found to be very high in gram negative enteric bacteria. In *E.coli* strains, 66% resistance was found to ampicillin, 16% to amoxicillin clavulanat, 28% to ampicillin/sulbactam, 14% to ceftazidime, 25% to ceftriaxone, 6% to nitrofurantoin, 6% to phosphomycine, 0% to imipenem, 29% to trimethoprim/sulphamethoxazol, 23% to gentamicine. ESBL positive Gram negative bacteria rate was 69/456 (15%). A total of 59 of them were *E.coli*, 10 of them were *Klebsiella* spp.

Table 1. Distribution of the microorganisms by gender							
	Male	Female	Total				
E. coli	127 (%43)†	165 (%57)	292 (%64)				
Klebsiella spp.	28 (%39)	44 (%61)	72 (%16)				
Proteus spp.	21 (%58)	15 (%42)	36 (%8)				
P.aeruginosa	4 (%30)	9 (%70)	13 (3)				
Enterococcus spp.	11 (%46)	13 (%54)	24 (5)				
S.aureus	4 (%44)	5 (%56)	9 (2)				
CNS	6 (%60)	4 (%40)	10 (2)				
Total	181 (%40)	275 (%60)	456 (%100)				

† growth number

CNS: Coagulase negative Staphylococcus

antibiotics			
	Enterococcus spp	S.aureus	CNS
	n=24	n=9	n=10
Penicillin	%46	%78	%81
Clindamycin	-	%11	%15
Eritromicin	%48	%44	%40
Vancomycin	%0	%0	%0
Cefoxitine	-	%44	%53
Linezolid	-	%6	%0
Gentamicin High Level	%41	-	-
TMP/SMX	-	%16	%8
Gentamicin	-	%16	%7
Fosfomycin	%20	-	-
Nitrofurantoin	%0	-	-
Ampicillin	%26	-	-

Table 2. Resistance rates of Gram positive microorganisms obtained in urine cultures against various antibiotics

TMP-SMX:Trimethoptim-sulfamethoxazole

Table 3. Resistance rates of Gram negative microorganisms obtained in urine cultures against various antibiotics								
	Escherichia coli	Klebsiella spp.	Proteus spp.	P.aeruginosa				
	n=292	n=72	n=36	n=13				
Ampicillin	193/292 %66	72/72 %100	26/36 %71	-				
Amoxycillin/clavulanat	e 47/292 %16	18/72 %25	5/36 %15	-				
Ampicillin/sulbactam	82/292 %28	28/72 %39	10/36 %28	-				
Phosphomicine	18/292 %6	16/72 %22	11/36 %31	-				
Gentamicin	67/292 %23	19/72 %26	8/36 %23	1/13 %8				
Imipenem	0/292 %0	9/72 %13	1/36 %3	0/13 %0				
Meropenem	0/292 %0	6/72 %8	0/36 %	0/13 %0				
Nitrofurantoin	18/292 %6	12/72 %17	13/36 %36	-				
Cefepime	-	-	-	3/13 %25				
Ceftazidime	41/292 %14	8/72 %11	6/36 %18	2/13 %16				
Ceftriaxone	73/292 %25	12/72 %16	0/36 %0	4/13 %33				
TMP-SMX	85/292 %29	17/72 %23	15/36 %41	-				
Piperacillin/Tazobacta	m -	-	-	1/13 %7				

TMP-SMX:Trimethoptim-sulfamethoxazol

Discussion

Urinary tract infections are frequent and important infectious diseases in children. 5-6% of infections in children evaluated because of fever are caused by UTIs (7). At the first step of UTI, there comes the colonization of periurethral region with enteric pathogens. These enteric bacteria start to hold on to mucosa thanks to their virulence properties and the process of inflammatory response comes out, there can be spread in the course of time and infection can spread towards bladder and kidney (8).VUR, neuromuscular dysfunction, mixion disorder, constipation, bladder neck stenosis, presence of catheter facilitate, familial and genetic predispositions facilitate UTI development (9). It is reported that there is also familial and genetic susceptibility (10-13). The possibility to develop severe complications increases in the presence of anatomic disorders and in repeating infections (14). In acute pyelonephritis cases, VUR was detected by 25-40% with the help of kidney scintigraphy (15, 16). The possibility to develop renal scar in these patients was reported to be 15%. The data in which renal scar development rate was detected as 37.1% in children with VUR and as 14.3% in children without VUR are the results of a newly made research in Turkey (17). Peru et al. did not detect a significant difference in VUR prevalence between three groups (acute pyelonefritis, afebrile UTI, afebrile recurrent UTI) a with the study they carried out in 642 pediatric patients with UTI.

While scar development triggers hypertension, chronic pyelonefritis paves the way for end-stage kidney failure. In order to be successful in the treatment of urinary tract infection and its complications, it is important that early diagnosis should be made and treatment for pathogenesis should be started. There are approaches stating the importance of early intervention of the first doctor for accurate diagnosis and treatment, as there are less aggressive approaches (18). The reason for this is that the possibility of renal parenchymal defects development is 3-15% in the first 2 years after being diagnosed UTI (19).

In the studies carried out on this subject, it is observed that acute phase renal involvement gets decreased but scar formations cannot be prevented through proper and correct antibiotic treatment started in the first 24 hours after the symptoms occur (20). Renal inflammatory changes were seen in 57% of 278 infants in this study. Renal defect was detected by 75% in patients for whom antibiotic treatment got started in the 4th day or later. This rate was 41% in the group for whom it got started within 24 hours since the start of fever. Again in the same study, acute inflammatory changes were reported to be encountered more commonly in the presence of high degree VUR. Jacobson et al. reported that among patients who had renal scarring from pyelonephritis during childhood, 23 percent developed hypertension and 10 percent developed end-stage renal disease (21).

Recently-made studies show that UTI is determined by 7% in children of 2-24 months of age and 8% in children of 2-18 years of age (22, 23). Occurrence rate varied widely depending on age, sex, and race. Also, UTI rate is determined to be lower in circumcised boys (1, 24, 25) considering the last data reported by CDC, 8% of girls and 2% of boys suffer UTI until the age of 7. One of the most important bacteria having been isolated is E.coli by 85% (24). In order to make the diagnosis according to the guides taken as a basis by CDC, it is doubtful that Leucocyte Esterase determined by urineanalysis is positive and more than five leucocyte is seen in each field that is the presence of pyuria, and significant reproduction in the culture test required in its follow up (at least 50,000 CFU s/mL of a single uropathogen from urine) is obligatory to make the exact diagnosis (24).

The guides updated about laboratory examinations to be performed with mid-stream urine samples and the usage of imaging methods in a correct manner and at a correct time are loadstars especially for repetitive UTI (22). In a study that included 199 urine culture, initial and late samples were compared. When using a cutoff value of at least 10.000 colony forming units/ml, late samples were superior to the initial ones in reducing contamination of urine cultures (26). Urine concentrations of interleukin-8 (IL-8) is a noninvasive marker for VUR in infants. IL-8 is produced by epithelial cells of the renal tract in response to inflammatory stimuli. Elevated urine IL-8 levels in VUR and renal scarring have already been reported to increase during acute UTI (27). Gökçe et al. was reported that children with VUR have a high urine IL-6 concentration, and children with RPS have a high urine IL-8 concentration (28).

The reports and collections carried out in this field indicate that there are similar bacterial reproduction rates for each geographical region but antibiotic susceptibility rates are changeable due to regional differences (29-32).

Urinary culture results of 524 pediatric patients were assessed in India, and reproduction was detected in 186 of them (35.4 %) (33). The rate of VUR was 18/186 in the cases with reproduction (9.7%) and 4/338 in the cases without reproduction (1.2%). 69% of the reproduction cases were boys and 31% of them were girls. *E.coli* is at the first rank with 68.3% among all Gram negative bacteria that reproduced, *Enterococcus faecalis* became the most common one among Gram positive bacteria with 65.3% (33).

In preschool and school children bactriuria incidence was 0.7-1.9% in girls and 0.02-0.04% in boys (34). After the first infection, there is the risk of recurrence of UTI in 20-30% of boys and 40-60% of girls (24). Febril stated that UTI is one of the most common severe bacterial infections in small children and its prevalence ranges in 4.1-7.5% (24). Although urinanalysis (leukocyte esterase, nitrite, white blood cells detection) foreshadows UTI at the beginning and its sensitivity is over 73%, the specificity of it cannot pass over 80% (22, 35). The isolation of the factor by having culture antibiogram and monitoring susceptibility results are necessary for correct preferences to be able to be made in treatment and prophylaxis.

It is reported that during neonatal and infancy periods, 79% of UTI is caused by *E.coli*, 7% by *Klebsiella* spp, 7% by *Pseudomonas* spp, 4% by *Proteus* spp (36). Although *E.coli* is the factor in the great majority of the infections in preschool and schooling periods, *Staphylococcus* spp is held

responsible together with *E.coli* in pre-puberty period. ESBL positive *E.coli* and *Klebsiella* spp are isolated in and increasing frequency in UTI (1, 36). In our study, ESBL positive Gram negative bacteria rate was determined to be 69/456 (15%) in compliance with the findings of Turkey and other countries (37-40).

Increased rates of *E.coli* resistance have made amoxicillin a less acceptable choice for treatment, and studies have found higher cure rates with trimethoprim/sulfamethoxazole (22). Other treatment options include amoxicillin/clavulanate and cephalosporins. Guidelines from the American Academy of Pediatrics recommend limiting fluoroquinolone therapy to patients with UTIs caused by multidrug-resistant, Gramnegative bacteria, especially *P.aeruginosa* (41). Ciprofloxacin is approved by the U.S. Food and Drug Administration for complicated UTIs and pyelonephritis attributable to *E. coli* in patients one to 17 years of age.

Low dose long-term antibiotic prophylaxis is indicated in children with VUR or other anatomic malformations having high risk in terms of renal scar development and in children having suffered two urinary tract infection attacks in six months but having no anatomic malformations (42). In a randomized controlled trial of children with mild to moderate vesicoureteral reflux, it was found that prophylactic antibiotics for 12 months following a febrile UTI did not reduce the risk of subsequent UTI (43).

Used prophylactic antibiotic should provide high urine concentration with low serum level and should have minimum effects on faecal flora and should be cheap and well tolerated. High prophylactic doses trigger antibiotic resistance. The medications effective in preventing repetitive urinary tract infection, which are used in prophylactic treatment, and in which resistance strains are low, become prominent as second- or third-generation cephalosporins, amoxicillin/clavulanate, or nitrofurantoin, trimethoprim/sulfametaksazol. Although the period of time of prophylactic antibiotic treatment is debatable, it is suggested that prophylactic antibiotic administration should be continued for 1-2 years in the cases with VUR, 3-6 months in cases with recurrent urinary tract infection but without anatomic malformation (44).

Besides, it is reported that single dose or single daily oral antibiotic treatments are less effective and thus are not suggested (45-47). There are studies defending that lowdose long-term antibiotic prophylaxis is proper in children with VUR or other anatomic malformations having high risk in renal scar development an in children having urinary tract infection more than 2 in 6 months but without anatomic malformation. Used prophylactic antibiotic should provide high urine concentration with low serum level and should have minimum effects on faecal flora and should be cheap and well tolerated (48). Because high prophylactic doses increase antibiotic resistance, it can cause harm rather than benefit.

Through the use of this, the study findings are promising about the fact that intravesical administrations to be performed with substances such as glycosaminoglycan (GAG), heparin can be preventive from UTI (49, 50). GAG layer prevents bacteria from binding surface epithelium cells. Also, GAGs are thought to play a role in excitement of immune system by promoting leukocyte activation and behavior in infected tissues (51). After the treatment, its preventive effect lasting for months can be explained with these functions of its. In a study in which intravesical heparin 40.000 U was given once a week for 6 weeks to 18 adult women with recurrent UTI history, the number of infections had 6 months ago and 6 months after the administration were recorded (52).

In conclusion, the recognition of risk factors for UTI in children may enable proper management of these patients and empirical antibiotic selection should be based on knowledge of the local prevalence of bacterial organisms and antibiotic sensitivities, because resistance patterns may vary in different regions.

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